

DETECTION OF PHYLOGENETIC FOOTPRINTS IN LARGE GENE CLUSTERS

S.J. Prohaska, C.Fried, C.Flamm¹, G.P. Wagner², P.F. Stadler

Lehrstuhl für Bioinformatik, Institut für Informatik, Universität Leipzig, Germany

¹Institut für Theoretische Chemie und Strukturbiologie, Universität Wien, Austria

²Dept. of Ecology and Evolutionary Biology, Yale University, New Haven CT.

Tel: ++49 341 149 5120

Fax: ++49 341 149 5119

Email: {sonja, claudia, xtof, studla}@bioinf.uni-leipzig.de

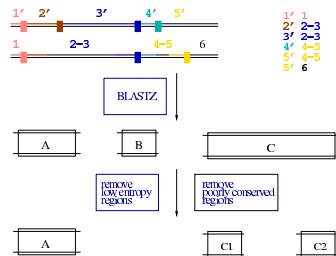
WorldWideWeb: <http://www.bioinf.uni-leipzig.de/>

Non-coding DNA in the genome of eukaryotes contains a large amount of functional regions for the regulation of genes expression. During evolution the **gene regulatory regions** are subject to stabilizing selection and therefore evolve much slower than adjoined non-functional DNA. These so-called **phylogenetic footprints** can be detected with the technique of **Phylogenetic Footprinting**. We present here a new method for **Phylogenetic Footprinting** based on comparison of the sequences surrounding orthologous genes in different species which handles a large set of sequences at once. Compared to the most recent program FootPrinter [1] our method does not require the phylogeny as input.

The program tracker is designed to survey large clusters without relying on a pre-supposed phylogeny.

The tracker approach

The program tracker is based upon **local alignments** of pairs of **orthologous intergenetic regions**. As blastz searches [5] with non stringent parameters can lead to the affiliation of alignments with **repetitive sequences** or **low conserved blocks** into the list, we have to remove the concerning stretches of these elements.



We use a **local entropy measures** to identify and to remove **repetitive sequences**.

$$H(k) = - \sum_a f_a(k) \log_2 f_a(k)$$

$$H_\tau(k) = - \sum_{a,b} f_{ab}^\tau(k) \log_2 f_{ab}^\tau(k)$$

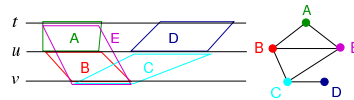
The measures are based on the nucleotide frequencies $f_a(k)$ and the joint frequencies $f_{ab}^\tau(k)$ of finding the nucleotides a and b separated by a distance τ along the chain. The values for $H(k)$ and $H_\tau(k)$ are low if the sequence **composition is restricted** and **highly repeated**.

The alignments consisting of a few highly conserved blocks separated by **long stretches of completely non-conserved sequences** are split at regions of low sequence identity.

To further construct **multiple alignments** based on the list of local matches we **combine overlapping alignments to clusters**.

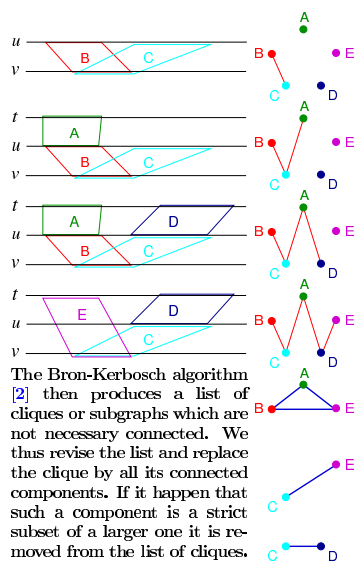
Alignments overlap and are clustered if they have at least one sequence from the same organism and the two sequences intervals from this organism do overlap.

A cluster is a graph Γ (overlap graph) with alignments as its vertices and edges for overlaps.



Using this definition we also get clusters that **cannot be represented by a common multiple alignment** (as given in the example above). This **incompatibility problem** is resolved by designating pairs of alignments as incompatible and building maximum consistent cliques.

For each cluster we obtain a list of incompatible alignments by following a path in the overlap graph that leads to **non-overlapping intervals in the same species**.

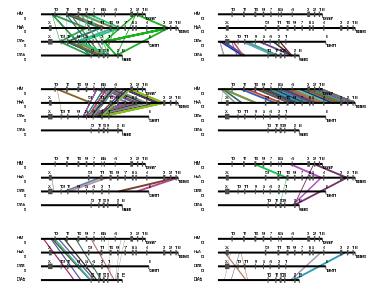
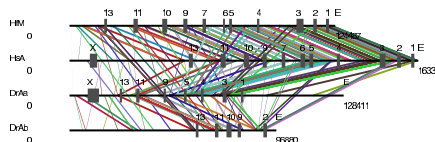


The Bron-Kerbosch algorithm [2] then produces a list of cliques or subgraphs which are not necessary connected. We thus revise the list and replace the clique by all its connected components. If it happen that such a component is a strict subset of a larger one it is removed from the list of cliques.

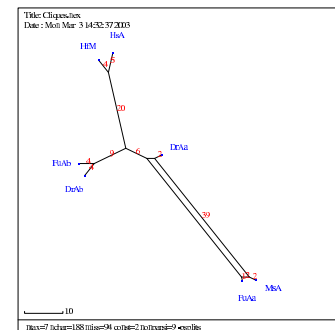
In the final step multiple alignments of all cliques are produced using a standard program such as DIALIGN.

Biological application

We applied tracker to the reevaluation of footprint analyses in the HoxA cluster sequences of shark (HfM), human (HsA), stripe bass (MsA), and the duplicated clusters of zebrafish (DrAa, DrAb) recently reported by Chiu et al. [3]. The purpose was to study the effect of **Hox cluster duplication** on the **pattern of regulatory sequences** in the non-coding region. Our main result is that the automatized procedure detects a more completes set of footprints.



Furthermore we extended the analysis to include both copies of Fugu HoxA clusters. The Buneman graph created using splitstree 3.1 shows the evolution of (co-linear) footprint cliques:



Statistical evaluation of the tracker output yields:

Cluster	Footprint loss		$\hat{\alpha}$
	data	model	
DrHoxAa	0.49	0.69	0.29
DrHoxAb	0.51	0.62	0.18
DrHoxA	0.49	0.66	0.26
TrHoxAa	0.45	0.58	0.22
TrHoxAb	0.21	0.40	0.48
TrHoxA	0.37	0.52	0.29

The observed rate of phylogenetic footprint loss is higher than predicted by the model introduced in [4]. It assumes that the **loss of phylogenetic footprint** can be explained entirely by the **loss of genes**. The deviation from the model given by $\hat{\alpha}$ is indicating for **additional non-structural causes of conservation loss**.

References

- [1] M. Blanchette, B. Schwikowski, and M. Tompa. Algorithms for phylogenetic footprinting. *J. Comp. Biol.* 9:211–223, 2002.
- [2] C. Bron and J. Kerbosch. Algorithm 457: Finding all cliques of an undirected graph. *CACM*, 16:575–577, 1973.
- [3] C.-h. Chiu, C. Amemiya, K. Dewar, C.-B. Kim, F. H. Ruddle, and G. P. Wagner. Molecular evolution of the HoxA cluster in the three major gnathostome lineages. *Proc. Natl. Acad. Sci. USA*, 99:5492–5497, 2002.
- [4] S. Prohaska, C. Fried, C. Flamm, G. Wagner, and P. F. Stadler. Surveying phylogenetic footprints in large gene clusters: Applications to Hox cluster duplications. *J. Mol. Biol.*, 2003. submitted; SFI preprint #03-02-011.
- [5] S. Schwartz, Z. Zhang, K. A. Fraser, A. Smith, C. Renner, J. Bouck, R. Gibbs, B. Hardison, and W. Miller. Pip2mer — a web server for aligning two genomic dna sequences. *Genome Research*, 4:577–586, 2000.

Printed by the Universitätsrechenzentrum Leipzig